ANSC (FSTC) 607 Physiology and Biochemistry of Muscle as a Food MOTOR INNERVATION AND MUSCLE CONTRACTION

I. Motor innervation of muscle

- A. Motor neuron
 - 1. Branched (can innervate many myofibers) \rightarrow terminal axons.
 - 2. Absolute terminal innervation ratio: one myofiber innervated by one terminal axon.
- B. The motor unit
 - 1. Each motor neuron innervates several muscle fibers within a muscle.
 - 2. Size of motor unit varies with muscle and fineness of movement.
 - a. There are 100 to 200 myofibers per motor neuron in larger muscles.
 - -- rat soleus, 200 fibers/neuron; rat gastrocnemius, 1,000 fibers/neuron
 - b. There are fewer in muscles such as ocular muscles.
- C. The neuromuscular junction
 - 1. Terminal axon
 - a. Vesicles (contain acetylcholine)
 - b. Presynaptic membrane
 - c. Synaptic cleft



Fig. 6.2. The fine structure of a vertebrate motor end-plate and nerve ending, seen in longitudinal section, as determined by light microscopy. *mf*, myofibril; *m.n.*, muscle nucleus; *ax*, axon; *tel*, Schwann cell (teloglia) nucleus; *sarc*, sarcoplasm; *my*, myelin sheath. (From Couteaux, 1960.)

- 2. Myofiber
 - a. Postsynaptic membrane *sarcolemma*
 - b. Synaptic clefts *increase surface area*







- D. Transmission of impulse across the synaptic cleft synaptic delay of the action potential
 - 1. Acetylcholine
 - a. End-plate potentials
 - b. Quantal nature of transmitter release each vesicle contains 10^3 to 10^4 molecules of acetylcholine
 - 2. Acetylcholinesterase
 - a. In synaptic cleft degrades acetylcholine
 - b. Stops transmission signal, contraction

II. Neurotransmitter release

- A. Neurotransmitter released from the presynaptic vesicles
- B. Stenin (myosin-like) is associated with vesicles and neurin (actin-like) is associated with the presynaptic membrane.
- C. Calcium influx causes stenin to fuse with neurin.
- D. This activates neurostenin ATPase; vesicles expel 1,000 to 10,000 molecules of acetycholine into cleft.





Electron micrograph of synaptosomal-enriched fraction from bovine brain. Neurin and stenin were enriched from this fraction. Puszkin et al. (1972) J. Neurochemistry 19:1319-1333

IV. Depolarization of the sarcolemma

- A. Depolarization of postsynaptic membrane
 - 1. Causes local depolarizaton (miniature end-plate potentials).
 - 2. Miniature end-plate potentials are caused by the quantal release of acetylcholine.
- B. Depolarization of sarcolemma
 - 1. Depolarization = summation of miniature end-plate potentials leads to action potential.
 - 2. Action potential spreads across sarcolemma.



3. Action potential reaches interior via the t-tubules.

VI. Generation of the resting membrane potential

A. Resting state (steady state)

requirements

- 1. Equimolarity
- 2. Electrical neutrality
- 3. Zero electrochemical gradient
- B. Basis for the resting membrane

potential

1. Ions responsible are primarily Na^+ and K^+ .

2. Factors influencing magnitude of

the action potential are primarily

the concentrations of Na^+ and K^+ .

- C. Calculation of the resting membrane potential
 - 1. Nernst equation: $E = (-RT)/zF ln[K_i]/K_0]$

Where E = potential difference across the membrane (usually in mV) R = gas constant

T = absolute temperature

Fig. 2-3. – Schematic representation of the distribution of major ions on either side of the membrane of a frog muscle cell. The column to the right of the ions indicates the concentration of each, in millimoles per liter. Pn⁻ is intracellular protein; the number in parentheses is the number of millequivalents of protein; i.e., the equivalent concentration of univalent ion. Note that intracellular fluid is rich in potassium (K⁺) and that the extracellular medium is high in so-dium (Na⁺) and chloride (Cl⁻).

Na† 117.5	Na ⁺ 10
κ ⁺ 2.5	к+ 140
CIT 120	CI ⁻ 2.5
	Pn ⁻ 97.5 (147.5)

F = Faraday's constant (# charges per mole ion) z = valence of ion

2. Modified Nernst equation

R and F = constants T (20°C) = 293 absolute z for K⁺ = 1 Convert *ln* to \log_{10} , so that: E (in mV) = -58 $\log_{10}[K_i]/[K_0]$

VI. The triad

A. Junction of the terminal cisternae of the sarcoplasmic reticulum and the transverse tubule (t-tubule)

B. Site of transmission of the action potential to the sarcoplasmic reticulum, via the t-tubule.





VII. Initiation of contraction

A. Action potential causes release of Ca⁺⁺ from the sarcoplasmic reticulum.

- Release of Ca⁺⁺ occurs in area of triad (at A band-to-I band interface in mammals).
- The dihydropyridine receptor (embedded in the T-tubule) is altered by the incoming action potential.
- 2. This causes the ryanodine receptor (underlying the Ttubule) to open, allowing calcium release from the sarcoplasmic reticulum.
- 3. Ca⁺⁺ follows its

electrochemical gradient into the sarcoplasm.

4. The sarcoplasmic

concentration of Ca++

increases from 10 nM to

10 µM (1,000-fold

increase).

B. Ca⁺⁺ binds to troponin C, which initiates contraction.



FIG. 1. Model of the triad junction. For simplicity, only the α_1 subunit of the dihydropyridine receptor and only one subunit of the ryanodine receptor homotetramer are shown. The ryanodine receptor is modeled after that of Takeshima *et al.* (17) although alternative models suggest as many as 12 transmembrane domains (18) and predict other regulatory domains within the foot region (19). *Question marks* indicate the speculative nature of the interaction between 94-kDa glycoprotein (triadin) and the ryanodine receptor as well as between triadin and calsequestrin.

VIII. Return to resting state

- A. Acetylcholine levels are reduced.
 - 1. Acetylcholine is repackaged into vesicles.
 - 2. Acetylcholine also can be degraded to acetate and choline by acetylcholinesterase.
- B. Sarcoplasmic Ca⁺⁺ resequestered in sarcoplasmic reticulum by:
 - 1. Ca⁺⁺-Mg⁺⁺-ATPase (active pumping)
 - 2. Calsequestrin
 - a. Binds 43 Ca⁺⁺/mol of protein.
 - b. Helps to concentrate Ca⁺⁺.
 - 3. High-affinity Ca⁺⁺-binding protein also helps to sequester Ca⁺⁺.

