A GRILIFE EXTENSION

Ammonia Emissions from Cattle Feeding Operations

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Issues and Emissions

Ammonia (NH₃) is a lighter-than-air, colorless gas with a recognizable pungent smell. It occurs naturally and is normally found in trace amounts in the atmosphere, where it is the dominant base, combining readily with acidic compounds. Ammonia is produced by the decomposition or fermentation of animal and plant matter containing nitrogen (N), including livestock manure, and is a source of the essential nutrient nitrogen for plants and animals. However, it is also classified as a hazardous substance by the US Environmental Protection Agency (EPA) due to concern about its potential to negatively affect air and water quality, and human and animal health.

Sources and emissions

Concentrated animal feeding operations (CAFOs) import feed ingredients that contain large quantities of nutrients such as nitrogen. Cattle retain a proportion of the nitrogen they consume, but approximately 70 to 90 percent is excreted in feces and urine (Cole et al., 2008). The breaking down of nitrogenous molecules

*Texas AgriLife Research, **USDA-Agricultural Research Service, ***Texas A&M AgriLife Extension Service in manure, such as urea and protein, produces ammonia. Urea in urine rapidly converts to ammonia and is a major ammonia source in manure, while microbes decompose more complex nitrogen-containing compounds, such as proteins, more slowly.

Historically, ammonia was considered a problem only within livestock buildings with inadequate ventilation or poor management.



Concentrated animal feeding operations import feed ingredients that contain large quantities of nitrogen. (Photo courtesy of S. Preece)

High ammonia levels negatively affect animal health and production and threaten the health of humans working inside. Correcting ventilation problems and periodically removing animal waste reduces ammonia levels within buildings, but these measures do not address the problem of ammonia emissions in the atmosphere from open-lot CAFOs.

Ammonia begins to volatilize (convert to a gas and be lost to the atmosphere) almost immediately after urea is excreted. The loss can continue as manure is handled, stored, or land-applied as fertilizer. Nitrogen is an essential plant nutrient and a primary component of fertilizer; nitrogen lost to the atmosphere from manure by ammonia volatilization is a loss of fertilizer value.

Ammonia in the atmosphere eventually returns to the Earth and is deposited as gas, particulates, or in precipitation onto surfaces such as soil or water. Ammonia deposition on nutrient-starved farmlands may be beneficial to crops; however, deposition in sensitive areas may be undesirable.

The complexity of biological and chemical processes, coupled with management decisions, complicates the understanding of ammonia emissions from livestock operations. Differences in livestock digestive systems, diets fed, feed and manure management systems, facility design, location, and weather are just a few of the factors that affect ammonia sources and emissions.

Environmental concerns

Undesirable ammonia deposition occurs when air currents transfer ammonia to sensitive land and water surfaces. Dry deposition occurs locally, and wet deposition occurs at longer distances from the source. Ammonia deposition can harm sensitive ecosystems when excessive nitrogen stimulates too much algae growth in surface waters, or weeds in fields or pastures. When algae growth dies, its decomposition consumes oxygen, resulting in hypoxia (low oxygen) in aquatic environments. For example, the hypoxic "dead zone" near the mouth of the Mississippi River is caused by excess nitrogen and phosphorus carried by the river into shallow coastal waters. This process of eutrophication is characterized by significant reductions in water quality; a disruption of natural processes; imbalances in plant, fish, and animal populations; and a decline of biodiversity.

Sensitive terrestrial ecosystems may experience excessive weedy plant growth, which outcompetes more desirable native species (Todd et al., 2004). Ammonia deposited in soil can undergo nitrification, which converts ammonia to nitrate. Nitrate is mobile in water and the nitrification reaction lowers (acidifies) the soil pH (Myrold, 2005). Forests in the humid eastern United States are especially susceptible to soil acidification, which can cause winter injury, loss of tree vigor, and the decline of desirable species.

The National Atmospheric Deposition Program (NADP, 2007) and the Clean Air Status and Trends Network (CASTNET) are excellent sources of long-term deposition data. Multiple monitoring stations located in strategic areas across the United States monitor and document wet and dry deposition of ammonium, nitrates, and other pollutants. Data from NADP and CASTNET are available online at http://nadp. sws.uiuc.edu/ and http://www.epa.gov/castnet/.

Human health concerns

Ammonia can significantly contribute to reduced air quality when it reacts with sulfur dioxide or nitrogen dioxide in the atmosphere to form aerosols. Aerosols, also known as particulate matter, are atmospheric particles classified by the EPA according to their aerodynamic diameter. Respirable aerosols are particles that can be inhaled deep into the lungs and have a mean aerodynamic diameter of less than 2.5 micrometers (PM_{2.5}). PM_{2.5} poses a threat to human health because it is associated with



Measuring ammonia emissions from concentrated animal feeding operations is difficult because ammonia quickly volatilizes and is dissipated by air currents. (Photo courtesy of S. Preece)

respiratory symptoms and diseases that lead to decreased lung function and, in severe cases, to premature death (EPA, 2009). Aerosols also affect cloud formation, alter the ozone layer, diminish irradiance, and reduce visibility in the air (Romanou et al., 2007; Chin et al., 2009).

Ammonia deposition can contaminate drinking water by increasing its nitrate concentration. This may occur by direct deposition onto water bodies or indirectly by leaching nitrogen from soils or by the erosion of nitrogen-laden soil particles into surface water.

Odor implications of ammonia are localized to regions in the vicinity of the CAFO. Ammonia is easily recognized by its smell, but is seldom associated with nuisance odor complaints near CAFOs any more than other manure constituents such as cresols, sulfides, or volatile fatty acids. Ammonia readily disperses from open-lot feedyards and dairies, which helps reduce its odor intensity to below human detection thresholds. Ammonia odors tend to be more noticeable inside animal barns than in open lots and are greater on or near CAFOs than at more distant off-site locations.

Measuring ammonia

Two categories of air quality measurements are commonly applied to ammonia at or near CAFOs: ambient concentrations and emission rates. Ambient concentrations are measurements of the ratio of ammonia to air in the atmosphere, usually measured in parts per million by volume (ppmv), parts per billion by volume (ppbv), or micrograms per cubic meter (µg/ m³). An accurate measurement of the atmospheric concentration in a large mass of dynamic, open air is difficult and requires special instrumentation and/or significant labor inputs.

Emission rates quantify ammonia flux from surfaces to the atmosphere and are reported in units of mass per unit area per unit time as in kilograms per square meter per day (kg/m²/day), and also in units of mass per unit animal per unit time such as kilograms per thousand head per year (kg/1000 hd/yr). Measuring ammonia emissions from non-point sources such as CAFOs is also difficult because once produced, ammonia quickly volatilizes and is dissipated by air currents. Quantifying ammonia flux from the feedyard surface to the atmosphere relies on direct measurement using fast-response instrumentation or with a flux model, which attempts to predict accurately the dispersion of gases and particulates through turbulent air. Emissions will vary depending on the type of surface (buildings, lagoons, pens) and the nature of processes at individual facilities.

Regulatory issues Federal reporting requirements (EPCRA)

Ammonia emission is regulated under the Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA) and the Emergency Planning and Community RightTo-Know Act (EPCRA). In December 2008, the EPA published a final rule that exempted CAFOs from reporting NH₃ emissions under CERCLA. However, under EPCRA [40 CFR §355 App A] CAFOs must report NH₃ emissions in excess of 45 kilograms (100 pounds) per day. Despite the challenges in accurately measuring ammonia emissions from CAFOs, an estimate of the lower and upper bounds can be calculated based upon animal headcounts and research-based figures for average emission rates per head. Non-compliance with the EPCRA NH₃ emission reporting requirements could result in fines of \$37,500 per day, criminal charges, and up to five years imprisonment.

Ammonia emissions may be indirectly addressed by federal and state regulations aimed at PM_{2.5} concentrations such as those in the National Air Quality Standards (NAAQS). Because ammonia is a precursor to $PM_{2.5}$, it may be necessary to reduce ammonia emissions in order to obtain a reduction in PM_{2.5} concentrations. Currently, there are few state regulations directed at ammonia emissions from animal agriculture. In 2003, California's Senate Bill 700 removed the reporting exemption from agricultural sources and, in 2006, Idaho put into force their Permit By Rule program requiring dairy farms with the capacity to produce more than 100 tons of ammonia annually to comply. Excepting Idaho and California, existing state agricultural ammonia regulations are aimed primarily at the distribution, storage, and land application of anhydrous ammonia fertilizer. However, states can directly address ammonia emissions in PM2.5 non-attainment areas in any case where ammonia is a significant contributor to PM_{2.5} concentrations. Some states base general air quality regulations on atmospheric concentrations and other states base them on actual emissions similar to those stipulated by EPCRA. However, atmospheric concentrations and ambient emissions of pollutants like ammonia are not well correlated. How these existing air quality regulations will

be applied to livestock ammonia sources in the future is unknown.

Ambient concentrations at cattle feedyards

Determining atmospheric concentrations of NH₃ requires sophisticated and expensive equipment, considerable labor, and much time. Measurements must be taken over large areas and during extended periods including all seasons to represent the large spatial and temporal variability. Other factors that must be reported include the animals, a detailed description of the facility, management practices, on-site weather, and sampling height. Data collected on atmospheric ammonia concentrations at CAFOs vary considerably, but tend to exhibit a 24-hour pattern, with daytime concentrations greater than those observed at night. Ammonia concentrations at cattle feedyards have rarely been observed over 3 ppm.

There are various methods for measuring atmospheric concentrations of ammonia, each with a unique set of advantages and disadvantages. Gas washing, denuders, and passive samplers provide average ammonia concentrations over relatively long periods of 1 to 4 hours. Gas washing is useful for calibration and standardization, but is labor-intensive. Fouriertransformed infrared (FTIR) spectroscopy, laser spectrometry, ultraviolet differential optical absorbance spectroscopy (UVDOAS), and chemiluminescence allow nearly real-time collection of measurements in relatively short periods of 5 seconds. Open-path lasers, UVDOAS, and FTIR have the added advantage of integrating measurements over distances from 50 to 500 meters. Because dust concentration in the vicinity of feedyards tends to be high, sampling for atmospheric ammonia requires special measures (such as installing Teflon filters preceding detectors or shortening measurement path lengths) to avoid errors.

Emission rates from cattle feedyards

An estimated 64 to 86 percent of total global anthropogenic (caused by human activity)

ammonia emissions come from CAFOs (Baum and Ham, 2009; EPA, 2008; Becker and Graves, 2004; Battye et al., 1994). Of the CAFO emissions, roughly 43 to 48 percent come from cattle operations (EPA, 2008; NRC, 2003; Battye et al., 1994). Figure 1 shows the relative contributions to ammonia emissions made by various US sources based on the National Emissions Inventory (EPA, 2008). This inventory considered ammonia emission factors and county-level livestock populations (beef cattle, dairy cattle, ducks, geese, horses, poultry, sheep, and swine) intentionally reared for the production of food, fiber, or other goods or for the use of their labor.



Figure 1. Estimated contributions of various US ammonia sources based on the National Emissions Inventory (EPA, 2008).

Extensive literature regarding ammonia emissions from swine and poultry facilities exists, but there is relatively little comprehensive research on large, open-lot beef cattle feedyards (Todd et al., 2008). Methods for estimating ammonia emissions from area sources such as feedyards include dispersion models, flux chambers, mass balance, micrometeorology, and wind tunnels (Hristov et al., 2011). The accuracy and applicability of these estimation methods varies greatly. For example, flux chambers and wind tunnels are appropriate for comparing treatments or assessing relative emission rates, but not for quantifying actual emissions (Cole et al., 2007; Paris et al., 2009; Parker et al., 2010). Dispersion models all rely on specific assumptions that are often challenged by the feedyard environment and can induce error in emission estimates (Flesch et al., 2005, 2007). Mass balance restraints are necessary to set an upper bound on emission estimates.

Calculating a total nitrogen balance for a facility involves determining the amount of nitrogen imported and exported from a feedyard and, assuming that unaccounted nitrogen is mostly ammonia, can provide reasonable estimates of ammonia emissions (Bierman et al., 1999; Farran et al., 2006; Cole and Todd, 2009). This is because the majority of gaseous nitrogen loss to the atmosphere is in the form of ammonia, as opposed to nitrous oxide (N₂O), nitrogen gas (N_2) , or nitrous oxides (NO_X) (Todd et al., 2005). To minimize errors, compare estimates obtained by multiple methods with calculations from a complete nutrient balance and local atmospheric concentration data. However, this approach is site-specific and impractical for regulatory monitoring at every livestock operation.

Micrometeorological methods such as eddy covariance (EC) and relaxed eddy accumulation (REA) are ideal for feedlots because they provide measurements of ammonia flux for large areas without disturbing the emitting surface. EC involves high frequency measurements using a fast-response analyzer, accounting for vertical air movements and the mixing ratio of ammonia in the air. REA is an adaptation of EC in which samples from air moving vertically are accumulated over time and analyzed with slowerresponse analyzers.

The most common method regulatory agencies use to estimate ammonia emissions

from CAFOs is to multiply a research-based emission factor by the number of animals on location. However, a single emission factor is not appropriate because ammonia emissions are affected by multiple, complex, and dynamic environmental variables. The National Research Council (NRC, 2003) has recommended a process-based modeling approach over the use of emission factors. Process-based models are based on the biological, chemical, and physical processes that contribute to emissions and take into account dynamic variables such as management practices, technologies, and weather conditions. Thus, they are applicable to a wide range of feedyard situations.

Research needs

Statistical, empirical, and process-based models estimate ammonia emissions from CAFOs. Statistical models are usually based on data collected from a particular location and provide estimates that may not be appropriate for a different site. Empirical models are commonly built from data collected under controlled conditions and predict well only when those particular conditions exist. Process-based (also known as mechanistic) models apply chemical and physical principles to a theoretical model of a real system, such as a CAFO. Their ability to predict ammonia emissions depends on how well the model represents real processes and the accuracy of important process factors used as inputs in the process-based model.

Many cross-disciplinary factors such as animal nutrition, environmental aspects, feedyard management strategies, and meteorological factors are considered in the construction of a process-based model. Process-based models of emissions from CAFOs often begin by describing the effects of diet and facility management on nutrient excretion by the animals. In the case of nitrogen, the various chemical forms, processes, and routes the nitrogenous molecules undergo as a feed constituent consumed and excreted by animals is described. Next, the nitrogenous manure constituents are accounted for and partitioned into several pools. Depending on the facility, these pools may include effluent lagoons, feces, manure stockpiles, pen surfaces, urine, and so forth. Finally, the chemical and physical transformations, transfer, and equilibria that occur during manure storage, handling, treatment, and export in each of the several cases are modeled. The model may then be used to predict ammonia emissions.

Models must consider atmospheric ammonia phases, which include gaseous ammonia (NH₃), fine particulate ammonia ((NH₄)₂SO₄ and NH₄NO₃), and liquid ammonia (NH₄OH) as clouds or fog. The transition between these three phases depends on other inconstant atmospheric constituents and the proportion of the phases relative to one another is also continually changing. Ammonia readily forms strong hydrogen bonds with water and will attach to many surfaces. Most materials exposed to air containing ammonia will absorb or adsorb ammonia compounds. In a CAFO environment, gaseous ammonia is prevalent and attaches to the airborne particulate matter emitted from the facility.

The dynamic nature of the atmosphere and its constituents results in significant variations in ammonia concentrations with respect to height



There is relatively little data on ammonia emission rates, emission factors, or flux rates from open-lot beef cattle facilities. (Photo courtesy of S. Preece)

above the ground, location, and time. Increasing the distance from the emission source can decrease ammonia concentrations, with the rate of decrease depending on other factors such as air temperature, relative humidity, or wind speed. Dry deposition rates close to the CAFO can also decrease with respect to distance and range widely depending on atmospheric conditions and emission rates.

Measuring emissions

It is difficult to measure ammonia emission rates from open-lot CAFOs. Ammonia tends to collect inside sampling instruments, adversely affecting measurement. Because open-lot CAFOs have lower ammonia concentrations than those typical of facilities with livestock housing, measuring emissions requires more sensitive instrumentation. There is relatively little data on ammonia emission rates, emission factors, or flux rates from open-lot beef cattle facilities.

Despite sampling challenges, changeability of ammonia concentrations, and scarcity of data, the average daily ammonia concentrations observed at several facilities by different researchers are consistent (Table 1).

When estimating ammonia emissions from open-lot beef cattle facilities, several compo-

nents of the CAFO system must be considered. Emission factors fail to account for the effects of particular components included in process-based models such as air and surface temperatures, animal age and diet, geographic location, time of year, and many others. So many variable and interactive system components must be considered that using a single emission factor is not adequate to predict ammonia emission rates (Hristov et al., 2011).

Processed-based models, which describe physical processes mathematically as opposed to statistically, are better suited to this task than emission factors. A single ammonia emission factor based primarily on European data proposed by the EPA (2005) is 13 kg/hd annually for feedlot cattle or 23 percent of the total amount of imported nitrogen. This EPA report also estimates the following nitrogen losses as ammonia: 1) stockpiles, 20 percent of nitrogen entering, 2) storage ponds, 43 percent, and 3) land application, 17 to 20 percent. Because European beef systems vary greatly from US systems, these values may not apply to US feedlot systems.

Studies conducted at North American feedyards using a variety of measurement methods observed a wide range of emission and flux (quantity per unit area per unit time) rates. Reported emission factors ranged between 18

STUDY	TIME	LOCATION	MEAN or RANGE
Hutchinson et al., 1982	April–July	Colorado	290–1,200
McGinn et al., 2003	May	Canada	66–503
	July		155–1,488
Todd et al., 2005	Summer	Texas	90-890
	Winter		10–250
Baek et al., 2006	Summer	Texas	908
	Winter		107
McGinn et al., 2007	June-October	Canada	46–1,730

Table 1. Ammonia concentrations (ug/m3) measured at several commercial open-lot beef cattle feedyards. (Adapted from Hristov et al., 2011.)

to 104 kg/hd annually, and flux rates ranged from 3.6 to 88 μ g/m²/s. Most studies also noted seasonal or 24-hour patterns in ammonia flux rates (Hristov et al., 2011). Reported losses from runoff holding ponds ranged from 3 to 70 percent of the nitrogen entering the pond. Compost piles, another source of ammonia loss on beef cattle feedyards, have been estimated to lose 10 to 45 percent of their initial nitrogen content (Hristov et al., 2011).

Abatement Measures

Ammonia abatement measures can be implemented at two different stages of livestock production. First-stage measures are applied preexcretion and include nutrition-based strategies to reduce the amount of nitrogen excreted in livestock manure. Second-stage measures occur post-excretion and use management strategies to reduce the amount of ammonia transferred

to the environment from the manure at agricultural operations.

Nutritional ammonia abatement methods

One means of reducing ammonia emissions from CAFOs is to reduce the amount of nitrogen excreted by the animals, especially the quantity excreted as urea in urine. Urinary pH can also affect ammonia emissions (Cole et al., 2008a). In some cases, it is possible to manipulate nutritional intake to reduce total nitrogen and urinary nitrogen excretion while continuing to meet the nutritional requirements and performance expectations of the animals. Based upon consistent observations among researchers over the past decade, annual ammonia losses from beef cattle feedyards tend to be approximately half of the nitrogen consumed by cattle, and summer emission rates are about twice those in winter (Todd et al., 2009) (Fig. 2).

There are a variety of ways to modify ration composition to reduce ammonia emissions by 20 to 50 percent with only small effects on animal performance (Cole et al., 2005, 2006a; Todd et al., 2006). Nutritional factors that can be manipulated include cation-anion balance (CAB), crude protein and/or degradable intake protein concentrations (including phase feeding), fat concentration, and fiber source and concentration, as well as some growth-promoting feed additives and implants. However, the large size of many CAFOs presents economic and logistic challenges when modifying diets or feeding practices. Modifications to diets, equipment,



Figure 2: Ammonia-N loss as a percentage of fed nitrogen from Great Plains beef cattle feedyards. Studies: (a) Todd and Cole, unpublished data, (b) Todd and Cole, unpublished data, (c) Todd et al., 2011, (d) Todd et al., 2008, (e) van Haarlem et al., 2008, (f) McGinn et al., 2007, (g) Flesch et al., 2007, (h) Harper et al., 2007, (i) Todd et al., 2005, (j) Todd et al., 2005, (k) Cole et al., 2006a, (l) Erickson and Klopfenstein, 2004, (m) Erickson et al., 2000, (n) Bierman et al., 1999.

or management practices may impose increased cost, labor, and time.

Crude protein

The concentration of protein in feed, as well as its ability to be degraded in the rumen, may affect the quantity and route of nitrogen excretion by beef cattle (Cole et al., 2005). Beef cattle consume dietary crude protein in two forms: degradable intake protein (DIP) and undegradable intake protein (UIP). Microbes process DIP in the rumen where it is either absorbed (normally as ammonia) or converted to microbial protein and nucleic acids. UIP escapes digestion in the rumen and passes to the intestine where it is digested and absorbed as amino acids (approximately 80 percent) or excreted (approximately 20 percent).

In general, as nitrogen consumption increases, urinary nitrogen excretion also increases. As the ratio of DIP to UIP increases, urinary nitrogen excretion also increases. Dietary changes must be made carefully and with consideration to unintended consequences. For example, in attempting to lower ammonia emissions, if the dietary protein intake is reduced below the animal's nutritional needs, the growth rate may be slowed, the animal will require more days on feed to reach market weight, and the cumulative ammonia emissions from a feedlot may actually increase. Making changes to decrease ammonia emissions may potentially result in increasing other undesirable emissions such as nitrous oxide.

In closed chamber laboratory (Cole et al., 2005) and artificial pen surface (Todd et al., 2006) experiments, decreasing the crude protein concentration of beef cattle finishing diets based upon steam-flaked corn from 13 to 11.5 percent decreased ammonia emissions by 30 to 44 percent. Ammonia fluxes from an artificial feedyard surface were reduced by 30 percent in summer, 52 percent in autumn, 29 percent in spring, and 0 percent in winter (Todd et al., 2006). The research team concluded that despite requirements to maintain cattle performance, reducing crude protein in beef cattle diets might be the most practical and cost-effective way to reduce ammonia emissions from feedyards. Another study by Todd et al. (2009) determined that feeding high concentrations (greater than 20 percent) of wet distillers grains, which are becoming increasingly available as a ration component, increased crude protein intake in beef cattle and resulted in increased ammonia emissions.

Phase feeding

As beef cattle mature, they require less dietary protein. Phase feeding involves adjusting nutrient intake over time to match the animal's changing needs. If protein is not progressively diminished in balance with the animals' nutritional requirements through the feeding period, potentially more nitrogen is excreted and more ammonia may be emitted from the facility (Cole et al., 2006a; Vasconcelso et al., 2009). Studies on cattle fed high-concentrate, steam-flaked, corn-based diets have suggested that a moderate reduction (approximately 1.5 percent) in dietary crude protein (CP) in the final 28 to 56 days of the feeding period may decrease ammonia emissions by as much as 25 percent with little adverse effect on animal performance (Cole et al., 2006a). Based on seven cooperative studies to determine the effect of crude protein on ammonia emissions and animal performance (Cole, 2006b), a reduction of dietary crude protein from 13 percent, which is optimal for growth, to 11.5 percent resulted in a 3.5 percent decrease in average daily gain and an approximate 30 percent reduction in ammonia emissions. In certain economic conditions, it may be practical to accomplish a significant reduction in ammonia emissions with a minimal effect on animal performance.

Distillers grains

Distillers grains have recently been introduced into beef cattle rations and may affect CAFO ammonia emissions. Research by Cole



Frequent pen cleaning may help capture nitrogen in the manure and decrease losses to the atmosphere. A Nebraska study revealed that cleaning pens once a month instead of after every five-month feeding period reduced apparent ammonia losses by 24 percent and increased the nitrogen content in the manure by 50 percent. (Photo courtesy of S. Preece)

et al. (2008b) reported that a 10 percent increase in distillers grains in rations based upon steamflaked corn increased manure production by approximately 10 percent. In rations based upon dry-rolled corn, the same increase in distillers grains resulted in a 0 to 7 percent increase in manure production. In both cases, the concentration of nitrogen in the manure was not affected. The combination of increased manure volume and steady nitrogen concentrations may result in potentially greater ammonia emissions. In a comparison of ammonia emissions at two feedyards, Todd et al. (2009) found that one feedyard feeding distillers grains averaged 149 grams of ammonia-N per head per day (NH₃-N head⁻¹ d⁻¹) over nine months, compared with 82 g NH₃-N head⁻¹ d⁻¹ at another feedyard feeding lower protein steam-flaked, corn-based diets.

Fiber

Manipulation of dietary fiber may also affect ammonia emissions from feedyards. In a study by Erickson et al. (2000), dietary fiber in the form of corn bran was increased in cattle finishing diets. During the winter-spring study period, nitrogen volatilization rates were decreased, but animal performance was adversely affected. In another study by Bierman et al. (1999), beef cattle were fed different diets containing alfalfa hay, corn silage, and wet corn gluten feed (WCGF). The researchers concluded that dietary fiber and carbohydrate source affected the way feed cattle digested and excreted feed, resulting in changes to the amount of nitrogen excreted. Nitrogen excretion was highest for cattle fed a ration based on WCGF, but these cattle also had the highest performance. Farran et al. (2006) manipulated alfalfa hay and WCGF in beef cattle diets and made similar observations. Increasing alfalfa hay or WCGF intake resulted in an increase in nitrogen intake, nitrogen excretion, nitrogen volatilization, and cattle performance. They further concluded that recovery of nitrogen in the manure and finished compost was also increased, especially in the case of WCGF, as a result of increased organic matter content in the manure.

Cation-anion balance (DCAB)

Ammonia emissions are inhibited in low-pH environments, and lowering dietary cation-anion balance (DCAB) can potentially lower the pH of cattle urine. Notwithstanding other factors, lowering the pH of cattle urine may potentially reduce CAFO ammonia emissions. However, Erickson and Klopfenstein (2010) noted no effect of DCAB on nitrogen volatilization losses. Lowering urine pH may have little effect on ammonia emissions because the pen surface of feedyard pens may have significant buffering properties that strongly resist pH changes, tending to maintain a pH of approximately 8 or higher (Cole et al., 2009). Furthermore, cattle performance may be reduced by low-DCAB diets (Cole and Greene, 2004).

Post-excretion ammonia abatement methods

Post-excretion ammonia abatement strategies, such as improving manure management, can reduce the rate of nitrogen volatilization and ammonia emissions. Animal health considerations in post-excretion methods are not as great a concern when compared to nutritional methods; however, some manure management strategies, such as pen scraping, can be beneficial for animal health. Manure contains nitrogen and phosphorus, both of which contribute to the value of manure as a fertilizer. Nitrogen volatilization can reduce the nitrogen to phosphorus ratio (N:P) to below most plant requirements, thereby reducing the fertilizer value of the manure and requiring a greater land application area to avoid excessive phosphorus applications. Reducing ammonia emission rates from manure will enhance the fertilizer value of manure and lower ammonia emissions. Besides manure management, manipulating other factors such as the pH and moisture content of soil and/or manure can also affect ammonia emissions (Cole et al., 2008a).

Urease inhibitors, zeolites, fats, and other pen surface amendments

Based upon laboratory studies, a number of compounds can potentially be applied to feedlot pen surfaces to reduce ammonia emissions from feedyard surfaces (Varel, 1997; Varel et al., 1999; Shi et al., 2001; Parker et al., 2005; Cole et al., 2007). Substances such as zeolites (a microporous, aluminosilicate mineral), fats, and urease inhibitors such as N-(n-butyl) thiophsphoric triamide, cyclohexylphosphoric triamide, and phenyl phospohorodiamidate may change manure properties such as pH, ammonia adsorption potential, or hydrolysis potential, which, in turn, affects ammonia emission rates.

Urease inhibitors work by slowing down or blocking the hydrolysis of urea (found in urine) by the enzyme urease (found in feces). However, urease inhibitors must continually be applied to manure because they rapidly degrade (Powers, 2002; Parker et al., 2005). Applying some compounds, such as fats, may be accomplished indirectly through dietary supplementation. Zeolites and urease inhibitors have been shown to decrease ammonia emissions when applied



Studies on cattle fed high-concentrate, steam-flaked, corn-based diets have suggested that a moderate reduction of about 1.5 percent in dietary crude protein in the final 28 to 56 days of the feeding period may decrease ammonia emissions by as much as 25 percent with little adverse effect on animal performance. (Photo courtesy of S. Preece)

as a surface amendment, but not when used as a dietary amendment (Varel 1997; Varel et al., 1999; Shi et al., 2001; Parker et al., 2005; Cole et al., 2007). Both dietary and surface amendments of fat appeared to decrease ammonia emissions (Cole et al., 2007). The dietary fat effect is likely because a proportion of fed fat is voided onto the feedyard surface after being excreted in undigested form by feedyard cattle. There were no significant effects on animal performance.

Lowering pH

One of the most important factors involved in ammonia emissions from surfaces is the pH of the emitting medium. In general, ammonia volatilization rates increase with pH. Lowering the pH of soil or manure can reduce ammonia emissions. With acidic conditions, given a constant temperature, more nitrogen will remain in the form of ammonium (NH₄⁺), thereby decreasing the amount of ammonia available to volatilize. A significant reduction in ammonia emissions has been observed with acidifying amendments such as aluminum sulfate (alum), ferrous sulfate, phosphoric acid, or calcium salts.

Maintaining the low pH can be challenging, however, because manure may have a strong buffering capacity, which results in the pH eventually returning to a more basic level and a resumption of ammonia emission. Strong acids are more cost-effective than weak acids or acidifying salts, but they are more hazardous and not suitable for use in agricultural environments (Ndegwa et al., 2008).

Manure harvesting, storage, and application

Frequent pen cleaning may help capture nitrogen in the manure by decreasing loss to the atmosphere. Research in Nebraska (Erickson and Klopfenstein, 2010) revealed that cleaning pens once per month, as opposed to once after every 166-day feeding period, reduced apparent ammonia nitrogen losses by 24 percent. The effectiveness of the monthly cleaning strategy varied seasonally, being less in winter. This may be due to the accumulation of nitrogen that occurs in the pen surface manure pack during the winter, apparently the result of decreased ammonia losses during the colder months (Cole et al., 2009). In addition, the amount of nitrogen collected in the manure was 50 percent greater from pens cleaned monthly.

Covering manure to reduce its exposure to elements such as rain, sun, and wind is very effective at reducing ammonia emissions from storage areas. When manure is land-applied, immediate incorporation or injection into the soil has been shown to significantly reduce ammonia losses when compared to broadcasting alone (Ndegwa et al., 2008).

References

Battye, R., W. Battye, C. Overcash, and S. Fudge. 1994. *Development and Selection of Ammonia Emission Factors, Final Report.* Washington DC, US Environmental Protection Agency, Office of Research and Development.

Baum, K. A. and J. M. Ham. 2009. "Adaptation of a Speciation Sampling Cartridge for Measuring Ammonia Flux from Cattle Feedlots Using Relaxed Eddy Accumulation." *Atmospheric Environment.* 43:(10) 1753–1759.

Becker, J. G. and R. E. Graves. 2004. *Ammonia Emissions and Animal Agriculture*. USDA Cooperative State Research, Education, and Extension System (CREES). Mid-Atlantic Water Quality Program.

Bierman, S., G. E. Erickson, T. J. Klopfenstein, R. A. Stock, and D. H. Shain. 1999. "Evaluation of Nitrogen and Organic Matter Balance in the Feedlot as Affected by Level and Source of Dietary fiber." *Journal of Animal Science*. 77:(7) 1645–1653.

Chin, M., R. Kahn, and S. Schwartz. 2009. Atmospheric Aerosol Properties and Climate Impacts: A Report by the US Climate Change Science Program and the Subcommittee on Global Change Research. Washington, DC. NASA. Cole, N. A. and L. W. Greene. 2004. "Nutrient Management for Livestock Feeding Operations: Challenges and Opportunities." In *California Animal Nutrition Conference Proceedings*, 82–104. Fresno, CA.

Cole, N. A., R. N. Clark, R. W. Todd, C. R. Richardson, A. Gueye, L. W. Greene, and K. McBride. 2005. "Influence of Dietary Crude Protein Concentration and Source on Potential Ammonia Emissions from Beef Cattle Manure." *Journal of Animal Science*. 83:(3) 722.

Cole, N. A., P. J. Defoor, M. L. Galyean, G. C. Duff, and J. F. Gleghorn. 2006a. "Effects of Phase-Feeding of Crude Protein on Performance, Carcass Characteristics, Serum Urea Nitrogen Concentrations, and Manure Nitrogen of Finishing Beef Steers." *Journal of Animal Science.* 84:(12) 3421.

Cole, N. A. 2006b. "Update on Recent Protein Research for Finishing Beef Cattle Fed Steam-Flaked Corn-Based Diets." In *Proceedings* of the 21st Annual Southwest Nutrition and Management Conference, 67-87. Tempe, AZ.

Cole, N. A., R. W. Todd, and D. B. Parker. 2007. "Use of Fat and Zeolite to Reduce Ammonia Emissions from Beef Cattle Feedyards." In *Proceedings of the International Symposium on Air Quality and Waste Management for Agriculture*, CD-ROM. ASABE (American Society of Agricultural and Biological Engineers) Paper No. 701P0907cd.

Cole, N. A., R. W. Todd, D. B. Parker, and M. B. Rhoades. 2007. "Challenges in Using Flux Chambers to Measure Ammonia Emissions from Simulated Feedlot Pen Surfaces and Retention Ponds." In *Proceedings of the International Symposium on Air Quality and Waste Management for Agriculture*, CD-ROM. ASABE Paper No. 701P0907cd.

Cole, N. A., R. Todd, B. Auvermann, and D. Parker. 2008a. "Auditing and Assessing Air Quality in Concentrated Feeding Operations." *The Professional Animal Scientist.* 24(1): 1–22. Cole, N. A., M. S. Brown, and J. C. Mac-Donald. 2008b. "Environmental Considerations of Feeding Bio-Fuel Co-Products." In "Symposium: ALPHARMA Beef Cattle Nutrition and Beef Species Joint Symposium: Producing Quality Beef in a Bio-Based Economy," supplement, *Journal of Animal Science*. 86:E–Suppl. 2.

Cole, N. A., and R. W. Todd. 2009. "Nitrogen and Phosphorus Balance of Beef Cattle Feedyards." In *Proceedings of the Texas Animal Manure Management Issues Conference*, 17–24. Round Rock, TX.

Cole, N. A., A. M. Mason, R. W. Todd, M. Rhoades, and D. B. Parker. 2009. "Chemical Composition of Pen Surface Layers of Beef Cattle Feedyards." *Professional Animal Scientist.* 25:(5) 541–552.

EPA. 2008. *National Emissions Inventory.* Environmental Protection Agency. Research Triangle Park, NC.

EPA. 2009. *Air Quality Index. A Guide to Air Quality and Your Health.* Environmental Protection Agency. Research Triangle Park, NC. EPA-456-F/09-002.

Erickson, G. E., C. T. Milton, and T. J. Klopfenstein. 2000. "Dietary Protein Effects on Nitrogen Excretion and Volatilization in Open-Dirt Feedlots." In *Proceedings of the Eighth International Symposium on Animal, Agricultural, and Food Processing Waste (ISAAF).* ASABE. St. Joseph, MI.

Erickson, G. and T. Klopfenstein. 2010. "Nutritional and Management Methods to Decrease Nitrogen Losses from Beef Feedlots." *Journal of Animal Science*. In press.

Farran, T. B., G. E. Erickson, T. J. Klopfenstein, C. N. Macken, and R. U. Lindquist. 2006. "Wet Corn Gluten Feed and Alfalfa Hay Levels in Dry-Rolled Corn Finishing Diets: Effects on Finishing Performance and Feedlot Nitrogen Mass Balance." *Journal of Animal Science.* 84:(5) 1205–1214. Flesch, T. K., J. D. Wilson, and L. A. Harper. 2005. "Deducing Ground-to-Air Emissions from Observed Trace Gas Concentrations: A Field Trial with Wind Disturbance." *Journal of Applied Meteorology*. 44:(4) 475–484.

Flesch, T. K., J. D. Wilson, L. A. Harper, R. W. Todd, and N. A. Cole. 2007. "Determining Ammonia Emissions from a Cattle Feedlot with an Inverse Dispersion Technique." *Agricultural and Forest Meteorology.* 144:(1–2) 139–155.

Harper, L. A., T. K. Flesch, R. W. Todd, and N. A. Cole. 2007. "Air Quality and Global-Change Gas Emissions in a Beef Feeding Operation." In *Agronomy Abstracts, 2004 Annual Meetings*. American Society of Agronomy. Seattle, MI.

Hristov, A., M. Hanigan, N. A. Cole, R. W. Todd, T. McAllister, T. P. Ndegwa, and C. A. Rotz. 2011. "Review: Ammonia Emissions from Dairy Farms and Beef Feedlots." *Canadian Journal of Animal Science*. 91(1): 1–35.

McGinn, S. M., H. H. Janzen, and T. Coates. 2003. "Atmospheric Ammonia, Volatile Fatty Acids, and Other Odorants Near Beef Feedlots." *Journal of Environmental Quality*. 34(4): 1173–1182.

McGinn, S. M., T. K. Flesch, B. P. Crenna, K. A. Beauchemin, and T. Coates. 2007. "Quantifying Ammonia Emissions from a Cattle Feedlot Using a Dispersion Model." *Journal of Environmental Quality*. 36:(6) 1585–1590.

Myrold, D. D. 2005. "Transformations of Nitrogen." In *Principles and Applications of Soil Microbiology*, 2nd ed., 333–372. D. M. Sylvia et al., ed. Pearson Prentice Hall, Upper Saddle River, NJ.

Ndegwa, P. M., A. N. Hristov, J. Arogo, and R. E. Sheffield. 2008. "A Review of Ammonia Emission Mitigation Techniques for Concentrated Animal Feeding Operations." *Biosystems Engineering*. 100(4): 453–469.

NRC. 2003. Air Emissions from Animal Feeding Operations: Current Knowledge, Future Needs. Ad Hoc Committee on Air Emissions from Animal Feeding Operations, Committee on Animal Nutrition, National Research Council. National Academies Press. Washington, DC.

Paris, C. S., D. B. Parker, N. A. Cole, R. W. Todd, E. A. Carraway, M. B. Rhoades, B. Baniya, and T. B. Brown. 2009. "Comparison of Ammonia Emissions Determined with Different Sampling Methods." In *Proceedings of the Texas Animal Manure Management Issues Conference*, 83–90. E. Jordan, ed. Round Rock, TX.

Parker, D. B., S. Pandrangi, L. W. Greene, L. K. Almas, N. A. Cole, M. B. Rhoades, and J. K. Koziel. 2005. "Rate and Frequency of Urease Inhibitor Application for Minimizing Ammonia Emissions from Beef Cattle Feedyards." *Transactions of the ASABE*. 48:(2) 787– 793.

Powers, W. 2002. "Emerging Air Quality Issues and the Impact on Animal Agriculture: Management and Nutritional Strategies." 49th Annual Maryland Nutrition Conference for Feed Manufacturers. Timonium, MD.

Rhoades, M. B., D. B. Parker, N. A. Cole, R. W. Todd, E. A. Caraway, B. W. Auvermann, D. R. Topliff, and G. L. Schuster. 2010. "Continuous Ammonia Emission Measurements from a Commercial Beef Feedyard in Texas." *Transactions of the ASABE*. 53 (6): 1823–1831.

Romanou, A., B. Liepert, G. A. Schmidt, W. B. Rossow, R. A. Ruedy, and Y. Zhang. 2007. "20th Century Changes in Surface Solar Irradiance in Simulations and Observations." *Geophysical Research Letters*. 34:L05713, doi:10.1029/2006GLO28356.

Shi, Y., D. B. Parker, N. A. Cole, B. W. Auvermann, and J. E. Melhorn. 2001. "Surface Amendments to Minimize Ammonia Emissions from Beef Cattle Feedlots." *Transactions of the ASABE*. 44:(3) 677–682.

Todd, R. W., W. Guo, B. A. Stewart, and C. Robinson. 2004. "Vegetation, Phosphorus, and Dust Gradients Downwind from a Cattle Feedyard." *Journal of Range Management*. 57:(3) 291–299. Todd, R. W., N. A. Cole, L. A. Harper, T. K. Flesch, and B. H. Baak. 2005. "Ammonia and Gaseous Nitrogen Emissions from a Commercial Beef Cattle Feedyard Estimated Using the Flux Gradient Method and N:P Ratio Analysis." In *Proceedings of the State of the Science: Animal Manure and Waste Management*, CD-ROM. San Antonio, TX

Todd, R. W., N. A. Cole, and R. N. Clark. 2006. "Reducing Crude Protein in Beef Cattle Diet Reduces Ammonia Emissions from Artificial Feedyard Surfaces." *Journal of Environmental Quality.* 35:(2) 404–411.

Todd, R. W., N. A. Cole, L. A. Harper, T. K. Flesch, and B. H. Baek. 2008. "Ammonia Emissions from a Beef Cattle Feedyard on the Southern High Plains." *Atmospheric Environment.* 42:(28) 6797–6805.

Todd, R. W., N. A. Cole, D. B. Parker, M. Rhoades, and K. Casey. 2009. "Effect of Feeding Distillers Grains on Dietary Crude Protein and Ammonia Emissions from Beef Cattle Feedyards." In *Proceedings of the Texas Animal Manure Management Issues Conference*, 83–90. E. Jordan, ed. Round Rock, TX. Todd, R. W., N. A. Cole, M. B. Rhoades, D. B. Parker, and K. D. Casey. 2011. "Daily, Monthly, Seasonal and Annual Ammonia Emissions from Southern High Plains Cattle Feedyards." *Journal of Environmental Quality.* 40(4): 1090–1095.

van Haarlem, R. P., R. L. Desjardins, Z. Gao, T. K. Flesch, and X. Li. 2008. "Methane and Ammonia Emissions from a Beef Feedlot in Western Canada for a Twelve-Day Period in the Fall." *Canadian Journal of Animal Science*. 88:(4) 641–649.

Varel, V. H. 1997. "Use of Urease Inhibitors to Control Nitrogen Loss from Livestock Waste." *Bioresource Technology*. 62:(1–2) 11–17.

Varel, V. H., J. A. Nienaber, and H. C. Freetly. 1999. "Conservation of Nitrogen in Cattle Feedlot Waste with Urease Inhibitors." *Journal of Animal Science*. 77:(5) 1162–1168.

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